



ELSEVIER

Archives of Medical Research ■ (2005) ■

**Archives
of Medical
Research**

ORIGINAL ARTICLE

Ozone Therapy Effects on Blood Biomarkers and Lung Function in Asthma

Frank A. Hernández Rosales, José L. Calunga Fernández, José Turrent Figueras,
Silvia Menéndez Cepero and Adonis Montenegro Perdomo

*Departamento de Biomedicina, Centro de Investigaciones del Ozono del Centro Nacional de Investigaciones Científicas,
Ciudad de la Habana, Cuba*

Received for publication February 18, 2005; accepted April 22, 2005 (ARCMED-D-00072).

Background. The relationship and behavior of serum immunoglobulin E (IgE) level, peripheral blood mononuclear cell (PBMC) human leukocyte antigen DR (HLA-DR) expression and erythrocyte glutathione antioxidant pathway in asthma patients treated with systemic ozone therapy, have not been studied before.

Methods. Asthma patients were treated about 1 year with three cycles (5 or 6 months each other) with three different ozone therapy protocols. Ozone major autohemotherapy (MAHT) was applied at doses of 4 and 8 mg, 15 sessions each cycle; and ozone rectal insufflations (RI) at a dose of 10 mg, 20 sessions each cycle. Serum IgE, HLA-DR expression in PBMC and biomarkers for antioxidant pathway were measured before and at the end of each cycle. Lung function and symptoms test were recorded at the beginning and after the third cycle.

Results. IgE and HLA-DR decreased with the three types of treatments, while increments in reduced glutathione, glutathione peroxidase, glutathione reductase and glutathione S-transferase were achieved with all treatments. Lung function and symptoms test were markedly improved. However, in all parameters the best response was obtained in the order: MAHT at 8 mg better than MAHT at 4 mg better than RI at 10 mg. Before ozone treatment, glutathione antioxidant parameters were under the normal reference values, suggesting the occurrence of oxidative stress associated with atopic asthma.

Conclusions. This study demonstrates the effectiveness of ozone therapy in reducing IgE and inflammatory mediators along with the induction of antioxidant elements. The study raises the role of systemic ozone therapy in atopic asthma by means of its immunomodulatory and oxidative stress regulation properties. © 2005 IMSS. Published by Elsevier Inc.

Key Words: Oxidative stress, HLA-DR, IgE, Antioxidant pathway, FEV1, FVC.

Introduction

Allergic asthma is characterized by airway hyperresponsiveness and airway inflammation. Population studies have shown that the majority of adults and children with well-documented asthma are atopic, and the prevalence of

asthma is greater in subjects with high serum immunoglobulin E (IgE) levels in a close relation with inflammatory cells (1–3). Examination of bronchoalveolar lavage (BAL) fluid cells and supernatants from allergic asthmatics has revealed the existence of Th2-like cytokine patterns (4,5) together with the IgE elevation (6). Increased IgE level in peripheral blood and bronchoalveolar tissue has been directly associated with atopic asthma pathology (7,8). Therefore, some therapeutic strategies today in atopic asthma are addressed to reduce IgE (9–11).

On the other hand, bronchial epithelial cells, lymphocytes and dendritic cells also play an active role in asthma.

Address reprint requests to: Frank A. Hernández Rosales, Dr.Sc., Departamento de Biomedicina, Centro de Investigaciones del Ozono del Centro Nacional de Investigaciones Científicas, A. Postal 6412, Ciudad de la Habana, Cuba; E-mail: frank.hernandez@cnic.edu.cu and frankozono@yahoo.com

After allergen activation they release increased amounts of inflammatory mediators (12–14) and express higher levels of human leukocyte antigen DR (HLA-DR) molecule (15–17), which have been identified as a marker for eosinophil activation and therefore implicated in the cellular network underlying inflammation in asthma.

Systemic ozone therapy has been proven to be effective to modulate immune system by inducing the production of cytokines from peripheral blood mononuclear cells (PBMC) (18,19) and to regulate the oxidative stress by inducing an increase of cellular antioxidant system (20–22).

Taking into account that Th2 cytokines are pivotal in regulating the allergic phenotype, the IgE response or the inflammatory cell-mediated function (23), we explore the hypothesis that systemic ozone therapy has a beneficial role in asthma by reducing IgE level and inflammatory mediators along with a positive influence on the antioxidant defense system.

Materials and Methods

Patients and Treatments

One hundred thirteen asthmatic patients (ages: 15–50 years) including both sexes were treated by ozone therapy. They were divided into three groups. Two groups were treated by ozone major autohemotherapy (MAHT) and the other one by ozone rectal insufflations (RI). All received three cycles of treatment. Each cycle comprised 15 sessions (five a week) for MAHT using an ozone dose of 4 mg (20 µg/mL per mL of blood and 200 mL of volume) in the first group, or 8 mg (40 µg/mL per mL of blood and 200 mL of volume) in the second one. MAHT was done by using sodium citrate 3.8% in blood transfusion glass flasks. Each cycle of the third group comprised 20 sessions using an ozone dose of 10 mg (50 µg/mL and 200 mL of gas volume). Time between cycles was 5 or 6 months in all groups. No other medication was given during the study.

All patients signed a specific consent to participate in the study and the Scientific and Ethical Committee from Ozone Research Center of Cuba approved the entire project.

Determinations

Blood samples were drawn from patients before and at the end of each cycle. Red blood cells, serum and PBMC were obtained by gentle centrifugation and Ficoll-Hypaque-dextran gradient. Total serum IgE quantification was done using a commercially available immunoenzymatic ELISA kit. For HLA-DR measurement, a flow cytometry study on lymphocyte subpopulation was done using the ior dr1 (IgG2a) anti HLA class II fluorescent (FITC) monoclonal antibody. Reduced glutathione (GSH) and glutathione reductase (GR) were measured in erythrocytes using the methods of Beutler (24). Erythrocyte glutathione peroxi-

dase (GPx) and glutathione-S-transferase (GST) activities were measured by the modification of Faraj et al. (25) to the method of Thomson (26) and by the method proposed by Habig and Jakoby (27), respectively.

Symptoms scores were recorded and respiratory function tests (forced expiratory volume at 1 sec [FEV1] and forced vital capacity [FVC]) were done using baseline spirometry tests before and at the end of each cycle.

Statistical Analysis

Analysis of the results was done using the values before starting the first cycle and those at the end of the third cycle of treatment. Results are expressed as mean ± SD. Comparison of matched samples was done by Wilcoxon test. A p value ≤0.05 was considered to be statistically significant.

Results

Patient Classification

Serum IgE and HLA-DR expression values before and after ozone therapy are reported in Table 1. A trend to diminish the values of both parameters was observed after ozone therapy because only HLA-DR with MAHT (8 mg) decreased statistically significantly. Before treatment, average IgE levels were 193 ± 73 IU/mL for MAHT (4 mg), 225 ± 87 IU/mL for MAHT (8 mg), and 196 ± 52 IU/mL for RI (10 mg), whereas HLA-DR averages were 32 ± 12% for MAHT (4 mg), 27 ± 12% for MAHT (8 mg), and 29 ± 12% for RI (10 mg). Very high abnormal values were not present in any of the three groups, indicating that all patients did not have the same asthma stage; therefore, they would not respond equally to the treatment.

Consequently, a distribution according to serum IgE level and PBMC HLA-DR expression was done (Table 2). Patients with very high figures in both parameters comprised only 43%. The rest of the patients had at least one of these parameters below the cut-off levels. We have established high cut-off levels of 250 IU/mL for IgE and 35% for HLA-DR; however, other authors assume cut-off

Table 1. Values of serum IgE and HLA-DR expression in PBMC before and after ozone therapy for the entire number of patients

Treatment	IgE (IU/mL)		HLA-DR (%)	
	Before	After	Before	After
MAHT (4 mg) ($n = 35$)	193 ± 73	194 ± 81	32 ± 12	29 ± 8
MAHT (8 mg) ($n = 41$)	225 ± 87	188 ± 78	27 ± 10	21 ± 2*
RI (10 mg) ($n = 37$)	196 ± 52	194 ± 74	29 ± 12	27 ± 9

MAHT, major autohemotherapy; RI, rectal insufflations; n , number of patients.

* $p \leq 0.05$.

177 **Table 2.** Subgroup classification according to high serum IgE
178 (>250 IU/mL) and high HLDR (>35%)

179 Subgroup	180 MAHT (4 mg)	181 MAHT (8 mg)	182 RI (10 mg)	183 Total (%)
184 High IgE + high HLA-DR	18	15	16	49 (43)
185 Normal IgE or HLA-DR	17	26	21	64 (57)
186 Total	35	41	37	113 (100)

187 MAHT, major autohemotherapy; RI, rectal insufflation.

188 levels between 121 and 150 for IgE (28–30), and <25% for
189 HLA-DR (16,31). This reason is because many patients are
190 in the normal subgroup in our study. We have established
191 high cut-off levels in order to be sure we are evaluating
192 a real atopic subgroup of asthmatic patients.

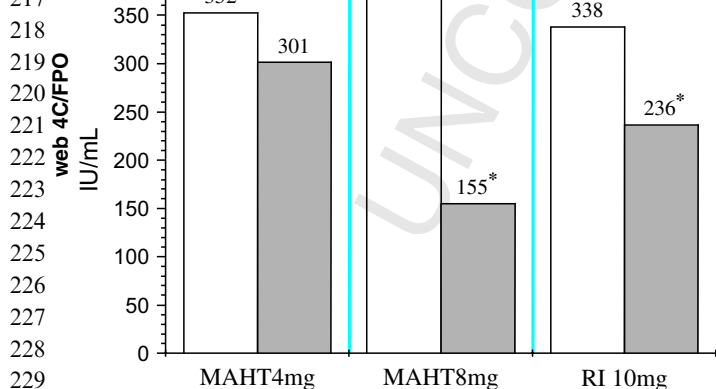
193 *Ozone Therapy Effects on IgE, HLA-DR and Glutathione*
194 *Antioxidant Pathway*

195 **Figures 1 and 2** show MAHT at a dose of 4 mg produced
196 a slight, non-statistically significant decrease in IgE level
197 (15%) and HLA-DR expression (10%) at the end of the
198 third cycle compared with the starting values. However,
199 MAHT at a dosage of 8 mg produced in IgE level (61%)
200 and HLA-DR expression (57%) statistically significant
201 decreases. Rectal insufflations also induced a significant
202 decrease in both parameters (30% for IgE and 40% for
203 HLA-DR).

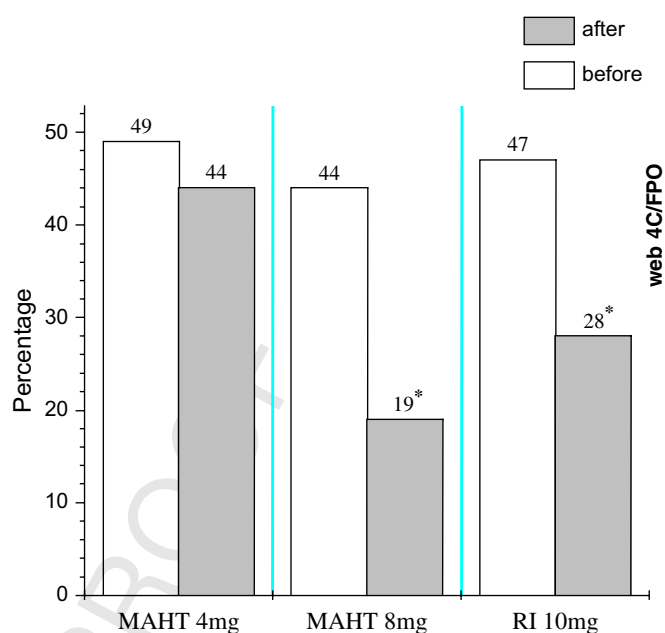
204 Results for glutathione antioxidant pathway are shown in
205 **Table 3**. GPx raised in a statistically significant manner
206 with MAHT (4 mg) at the end of the third cycle of ozone
207 treatment (8.90 vs. 11.26 IU/gHb). However, ozone therapy
208 with MAHT (8 mg) produced significantly higher values in
209 all of the antioxidant biomarkers (GSH = 1.78 vs. 2.86
210 μmol/gHb; GPx = 7.56 vs. 14.21 IU/gHb; GR = 4.50 vs.
211 6.02 IU/gHb; GST = 5.84 vs. 9.99 IU/gHb). Treatment
212 with RI (10 mg) stimulated remarkable increase in GPx
213 (8.35 vs. 11.50 IU/gHb) and GST (7.01 vs. 10.08 IU/gHb)
214 enzymes.

215 The changes that occurred in lung function are displayed
216 in **Table 4**. No statistical changes were observed in any of
217 the pulmonary function parameters with MAHT (4 mg).
218 Nevertheless, a positive tendency was observed in the three
219 parameters. MAHT-8 mg showed a significantly recovery
220 of FVC (2.34 vs. 2.99 L) and both forms of FEV₁ (1.46 vs.
221 2.20 L and 46.00 vs. 73.25%). A statistically significant
222 increase was also observed for both forms of FEV₁ (1.59 vs.
223 1.81 L and 50.53 vs. 59.63%) when RI (10 mg) was
224 applied. FVC was not statistically significantly increased.

225 Clinical improvement from episodic dyspnea, wheezing
226 and medication was better with MAHT (8 mg) than with RI
227 (10 mg), and better than MAHT (4 mg) (data not shown).



228 **Figure 1.** IgE values before first and after third cycle of ozone therapy in
229 patients with real atopic asthma. **p* < 0.05.



230 **Figure 2.** HLA-DR values before first and after third cycle of ozone
231 therapy in patients with real atopic asthma. **p* < 0.05.

232 6.02 IU/gHb; GST = 5.84 vs. 9.99 IU/gHb). Treatment
233 with RI (10 mg) stimulated remarkable increase in GPx
234 (8.35 vs. 11.50 IU/gHb) and GST (7.01 vs. 10.08 IU/gHb)
235 enzymes.

236 The changes that occurred in lung function are displayed
237 in **Table 4**. No statistical changes were observed in any of
238 the pulmonary function parameters with MAHT (4 mg).
239 Nevertheless, a positive tendency was observed in the three
240 parameters. MAHT-8 mg showed a significantly recovery
241 of FVC (2.34 vs. 2.99 L) and both forms of FEV₁ (1.46 vs.
242 2.20 L and 46.00 vs. 73.25%). A statistically significant
243 increase was also observed for both forms of FEV₁ (1.59 vs.
244 1.81 L and 50.53 vs. 59.63%) when RI (10 mg) was
245 applied. FVC was not statistically significantly increased.

246 Clinical improvement from episodic dyspnea, wheezing
247 and medication was better with MAHT (8 mg) than with RI
248 (10 mg), and better than MAHT (4 mg) (data not shown).

249 **Discussion**

250 In our opinion, airway hyperresponsiveness is closely
251 related to the redox and immunological status of asthma
252 patients; therefore, systemic ozone therapy with its
253 properties for oxidative stress regulation and modulation
254 of the immune system can be viewed as an efficient therapy
255 for atopic asthma.

256 In the present study we observed that the three groups of
257 ozone therapy treatments produced remarkable decreases in
258 IgE level and HLA-DR expression. Considering the
259 extension effect on each group, the results suggest that
260 both parameters are closely related because their responses

Table 3. Glutathione antioxidant pathway values before and after three cycles of ozone therapy in real atopic asthma patients

	MAHT (4 mg)		MAHT (8 mg)		RI (10 mg)	
	Before	After	Before	After	Before	After
GSH	1.65 ± 0.14	1.75 ± 0.39	1.78 ± 0.22	2.86 ± 0.34*	1.80 ± 0.10	1.93 ± 0.11
GPx	8.90 ± 1.11	11.26 ± 0.8*	7.56 ± 1.40	14.21 ± 1.98*	8.35 ± 0.98	11.50 ± 1.2*
GR	3.25 ± 0.80	3.46 ± 0.24	4.50 ± 0.71	6.02 ± 0.20*	3.89 ± 0.23	3.91 ± 0.4
GST	6.55 ± 0.35	6.22 ± 0.95	5.84 ± 1.21	9.99 ± 0.80*	7.01 ± 1.01	10.08 ± 1.1*

Values for GSH are expressed in $\mu\text{mol/g Hb}$, and for GPx, GR, and GST are expressed in IU/g.Hb.

* $p < 0.05$.

were apparently similar. Another conclusion drawn from these results is that ozone therapy effect is dependent on the applied route and ozone concentration.

It has been suggested that reactive oxygen species (ROS) play an important role in the pathogenesis of airway inflammatory diseases (32,33). In asthma patients, a correlation has been found between the degree of bronchial responsiveness to methacholine and the production of superoxide anion by peripheral polymorphonuclear leukocytes (34,35). Katsumata et al. reported that ROS could induce bronchoconstriction and airway hyperresponsiveness in anesthetized cats (36). On the other hand, glutathione and antioxidant enzymes have been found to protect the lung against ROS toxicity (37,38). These findings demonstrate that ROS exert a direct effect in mediating bronchial responsiveness and asthma. Because ozone therapy has the property to regulate the oxidative stress caused by ROS (20–22), we tried to determine if there is some relationship between asthma immunological mediators and antioxidant defense systems. One of these antioxidant defense systems is the glutathione antioxidant pathway, which includes the enzymes GPx, GST and GR, together with GSH metabolite.

In our study we found the three groups of patients had values under normal range for GSH ($< 2.00 \mu\text{mol/gHb}$) and GPx ($< 11.62 \text{ IU/gHb}$) before ozone treatment (Table 3), suggesting the existence of oxidative stress in these patients. This is evidence for the relationship between high IgE level and the oxidative stress occurrence. After ozone treatment MAHT at a dose of 4 mg induced a significant increase in GPx enzyme activity (approximately 130%), with non-statistically significant changes in the rest of parameters. MAHT at a dose of 8 mg influenced positively all the components of the glutathione antioxidant pathway; the major increase being in the GPx (approximately 190%), followed by GST (approximately 170%), GSH (approximately 160%), and finally GR (approximately 140%). RI at a dose of 10 mg produced significant enhancement in GPx (138%) and GST (144%) enzymes, and no significant changes in GSH and GR parameters. It is evident that MAHT at ozone dose of 8 mg induced the most powerful response in stimulating the glutathione antioxidant path-

way, followed by the response induced by RI at ozone dose of 10 mg. MAHT at 4 mg produced a mild response. Thus, the effect on glutathione antioxidant pathway was also dose and route dependent. This behavior was inversely similar to that seen for IgE and HLA-DR. For that reason, we assumed in these patients there was a close relationship between the asthma immunological mediators and the antioxidant system. It is important to point out that all groups of patients remarkably improved their oxidative stress condition. It has been proven that antioxidant therapy inhibits airway inflammation and hyperresponsiveness in a mouse model of asthma, suggesting that this may be useful as adjuvant therapy for bronchial asthma (39).

Marked improvement in objective function tests was achieved after three cycles of ozone therapy (Table 4). There were highly significant increments in the mean from FVC and FEV₁ with MAHT (8 mg). However, patients with MAHT (4 mg) did not have statistically significant increments of these function tests although a trend to elevate these values is observed. Patients with RI (10 mg) had a remarkable increase in FEV₁ but not in FVC. Comparison among the three groups of patients shows that the highest increment for FEV₁ was obtained with MAHT (8 mg) followed by RI (10 mg) and MAHT (4 mg) in

Table 4. Effect of ozone therapy on pulmonary function tests before and after three cycles of treatment

	Before	After
MAHT (4 mg)		
FVC (L)	2.39 ± 0.39	2.63 ± 0.56
FEV ₁ (L)	1.56 ± 0.33	1.79 ± 0.53
FEV ₁ (%)	49.60 ± 14.68	57.60 ± 20.36
MAHT (8 mg)		
FVC (L)	2.34 ± 0.28	2.99 ± 0.60**
FEV ₁ (L)	1.46 ± 0.20	2.20 ± 0.57***
FEV ₁ (%)	46.00 ± 8.10	73.25 ± 19.70***
RI (10 mg)		
FVC (L)	2.44 ± 0.45	2.59 ± 0.56
FEV ₁ (L)	1.59 ± 0.39	1.81 ± 0.52*
FEV ₁ (%)	50.53 ± 13.58	59.63 ± 20.46*

* $p < 0.05$; ** $p < 0.02$; *** $p < 0.001$.

397 agreement with the behavior for the responses of IgE,
398 HLA-DR and antioxidant biomarkers.

399 There was also marked improvement in clinical
400 symptoms. Clinical amelioration achieved in episodic
401 dyspnea, wheezing and medication was reported with the
402 same behavior pattern observed for lung function tests.
403 Therefore, the present data corroborate the beneficial
404 effects of ozone therapy in asthma patients.

405 In a previous paper we have demonstrated MAHT at 8
406 mg induces a significant reduction in serum IgE and HLA-
407 DR expression in PBMC, together with a modulation in
408 CD3⁺, CD4⁺ and CD8⁺ lymphocyte subpopulations (40).
409 In our present study, we found similar results for IgE and
410 HLA-DR together with remarkable induction of glutathione
411 antioxidant system and improvement in function and
412 symptom tests using MAHT at an ozone dose of 8 mg
413 and 4 mg and RI at 10 mg. Essentially the agreement in the
414 response pattern for all parameters suggests that ozone
415 therapy modulated the immune system for IgE production
416 along with the induction in antioxidant system. These
417 events allowed the improvement in the asthma state.

418 The mechanism for immune system modulation is not
419 yet totally proven. We can indirectly assume ozone therapy
420 influenced synthesis and release of cytokines (18,19) in
421 such a way that a change from Th2 to Th1 (IL-2, IFN γ)
422 cytokine pattern occurs, or ozone therapy acted as Th2
423 cytokine inhibitor. The change in cytokine pattern could be
424 the reason for the reduction in the release and expression of
425 asthma mediators (IgE, HLA-DR,) (41,42). Changes from
426 Th2 to Th1 cytokine pattern have been demonstrated to
427 counteract the airway hyperresponsiveness and bronchial
428 inflammation (43,44). In the presence of IL-4 and IL-13, the
429 B cell undergoes class switching to produce IgE (45),
430 which can induce the release of pro-inflammatory cytokines
431 including the expression of HLA-DR. It is important to
432 remark that the selection of ozone concentration is
433 a fundamental aspect in cytokine release (46,47). Our
434 results are in accordance with this concept because our best
435 results were in the proposed ozone concentration range for
436 cytokine release. Combination of the expressed actions may
437 be the possible mechanism by which ozone therapy has its
438 beneficial effect on asthmatic patients. However, experi-
439 mental evidence of cytokine type released after ozone
440 therapy application is needed. This could be our next
441 objective.

442 In summary, our study corroborates the outstanding role
443 of IgE and its direct relationship in the induction of
444 oxidative stress and subsequent bronchial inflammation in
445 allergenic asthma patients. Moreover, we have proven the
446 hypothesis that in these patients systemic ozone therapy
447 reduces IgE level and PBMC HLA-DR expression along
448 with antioxidant system induction. Thus, ozone therapy can
449 be seen as a new therapeutic or adjuvant approach for
450 atopic asthma, thanks to its immunomodulation and
451 oxidative stress regulation properties.

References

1. Burrows B, Martinez FD, Halonen M. Association of asthma with serum IgE levels and skin-test reactivity to allergens. *N Engl J Med* 1989;320:271–277. 452
2. Tuñon de Lara M. Immunoglobulines E et cellules de l'inflammation. *Rev Mal Resp* 1996;13:27–32. 453
3. Sunyer J, Anto JM, Castellsagué J, Soriano JB, Roca J. Total serum IgE is associated with asthma independently of specific IgE levels. *Eur Respir J* 1996;9:1880–1884. 454
4. Robinson DS, Hamid Q, Ying S, Tsicopoulos A, Barkans J, Bentley AM, Corrigan C, Durham SR, Kay AB. Predominant Th2-like bronchoalveolar T-lymphocyte population in atopic asthma. *N Engl J Med* 1992;326:298–304. 455
5. Taube C, Dakhama A, Gelfan EW. Insights into the pathogenesis of asthma utilizing murine models. *Int Arch Allergy Immunol* 2004;135:173–186. 456
6. Magnan A, Romanet S, Vervloet D. Asthma and allergy. *Rev Prat* 2001;15:511–516. 457
7. Maddox L, Schwartz DA. The pathophysiology of asthma. *Annu Rev Med* 2002;53:477–498. 458
8. Maddox L, Schwartz DA. The pathophysiology of asthma. *Annu Rev Med* 2002;53:477–498. 459
9. Brownell J, Casale TB. Anti-IgE therapy. *Immunol Allergy Clin North Am* 2004;24:551–568. 460
10. D'Amato G. Treating atopic asthma with the anti-IgE monoclonal antibody. *Monaldi Arch Chest Dis* 2002;57:117–119. 461
11. Stelmach I, Grzelewski T, Majak J, Bobrowska M, Jerzynska J, Kuna P. The effect of triamcilon, montelukast and formoterol on serum levels of IL-4 IgE and clinical parameters in children with asthma. *Pol Merkuriuz Lek* 2001;11:247–251. 462
12. Poston R, Litchfield P, Chanez P, Lacoste JY, Lee TK, Bousquet J. Immunohistochemical characterization of the cellular infiltration of asthmatic bronchi. *Am Rev Respir Dis* 1992;145:918–921. 463
13. Djukanovic R, Roche WR, Wilson JW, Besley CRW, Twentyman OP, Howarth PH, Holgate ST. State of the art: mucosal inflammation in asthma. *Am Rev Respir Dis* 1990;142:434–437. 464
14. Campbell AM, Chanez P, Vignola AM, Bousquet J, Couret I, Michel FB, Godard PH. Functional characteristic of bronchial epithelium obtained by brushing in from asthmatic and normal subjects. *Am Rev Respir Dis* 1993;147:529–534. 465
15. Vignola AM, Campbell AM, Chanez P, Bousquet J, Lacoste PP, Michel FB, Godard PH. HLA-DR and ICAM-1 expression on bronchial epithelial cells in asthma and chronic bronchitis. *Am Rev Respir Dis* 1993;148:689–694. 466
16. Kim YK, Oh HB, Oh SY, Cho SH, Kim YY, Min KU. HLA-DRB1*07 may have a susceptibility and DRB1*04 a protective effect upon the development of a sensitization to house mite *Dermatophagoides pteronyssinus*. *Clin Exp Allergy* 2001;31:110–115. 467
17. Jahnsen FL, Moloney ED, Hogan T, Upham JW, Burke CM, Holt PG. Rapid dendritic cell recruitment to the bronchial mucosa of patients with atopic asthma in response to local allergen challenge. *Thorax* 2001;56:823–826. 468
18. Bocci V. Ozone: a mixed blessing. New mechanisms of the action of ozone on blood cells make ozonated major autohemotherapy a rational approach. *Res Compl Med* 1996;3:25–33. 469
19. Bocci V, Luzzi E, Corradeschi E, Paulesu L, Rossi R, Cardaioli E, Di Simplicio P. Studies on the biological effects of ozone: 4 cytokine production and glutathione levels in human erythrocytes. *J Biol Regul Homeost Agents* 1993;7:133–138. 470
20. Bocci V. Does ozone therapy normalize the cellular redox balance? *Med Hypot* 1996;46:150–154. 471
21. Hernández F, Menéndez S, Wong R. Decrease of blood cholesterol and stimulation of antioxidative response in cardiopathy patients treated with endovenous ozone therapy. *Free Radic Biol Med* 1995;19:115–119. 472

[Q1]

452
453
454
455
456
457
458
459
460
461
462
463
464
465
466
467
468
469
470
471
472
473
474
475
476
477
478
479
480
481
482
483
484
485
486
487
488
489
490
491
492
493
494
495
496
497
498
499
500
501
502
503
504
505
506

- 507 22. Hernández F, Menéndez S, Gómez M, Eng L. Efecto de la
508 ozonoterapia intravascular sobre el sistema de la glutatión peroxidasa.
509 Rev CENIC Cienc Biol 1989;20:37-40.
- 510 23. Maeda S, Yanagihara T. Inflammatory cytokines (IL-4, IL-5 and IL-
511 13). Nippon Rinsho 2001;59:1894-1899.
- 512 24. Beutler E. Red Cell Metabolism: A Manual of Biochemical Methods.
513 New York: Grune & Stratton;1971. pp. 64-103.
- 514 25. Faraji B, Kang HK, Valentine JL. Methods compared for determining
515 glutathione peroxidase activity in blood. Clin Chem 1987;33:539-543.
- 516 26. Thomson CD, Rea HM, Doesburg VM, Robinson MF. Selenium
517 concentration and glutathione peroxidase activities in whole blood of
518 New Zealand residents. Br J Nutr 1977;37:457-460.
- 519 27. Habig WH, Jakoby WB. Methods in Enzymology, Vol. 77. New York:
520 Academic Press;1981. pp. 398-405.
- 521 28. Okudaira H, Hongo O, Ogita T, Haida M, Yamauchi N, Miyamoto T.
522 Serum IgE and IgE antibody levels in patients with bronchial asthma,
523 atopic dermatitis, eosinophilic granulomas of the soft tissue (Kimura's
524 disease) and other diseases. Ann Allergy 1983;20:50-54.
- 525 29. Zetterstrom O, Johanson SGO. IgE concentrations measured by
526 PRIST in serum and healthy adults and in patients with respiratory
527 allergy. Allergy 1981;36:537-542.
- 528 30. Mori A, Suko M, Nishizaki Y, Kaminuma O, Kobayashi S,
529 Matsuzaki G, Yamamoto K, Ito K, Tsuruoka N, Okudaira H. IL-5
530 production by CD4⁺ T cells on asthmatic patients is suppressed by
531 glucocorticoids and the immunosuppressants FK506 and cyclosporin
532 A. Int Immunol 1995;7:449-457.
- 533 31. Tsai JJ, Ma JK, Wang TF, Wang SR, Kao DH. The modulatory effect
534 of tetrandine on the CD23, CD25 and HLA-DR expression and
535 cytokine production in different groups of asthmatic patients. Int Arch
536 Allergy Immunol 1995;108:183-188.
- 537 32. Juniper ET, Firth PA, Hargreave FE. Airway responsiveness to
538 histamine and methacholine: relationship to minimum treatment to
539 control symptoms of asthma. Thorax 1981;36:575-579.
- 540 33. Kanazawa H, Kurihara N, Hirata K, Takeda T. The role of free
541 radicals in airway obstruction in asthmatic patients. Chest 1991;100:
1319-1325.
- 542 34. Metzger S, Goldberg B, Lad P, Easton J. Superoxide generation and its
543 modulation by adenosine in the neutrophils of subjects with asthma.
544 J Allergy Clin Immunol 1989;83:960-963.
- 545 35. Kamoi H, Kurihara N, Fujiwara H, Hirata K, Takeda T. The macrolide
546 antibacterial Roxithromycin reduces bronchial hyperresponsiveness
547 and superoxide anion production by polymorphonuclear leukocytes in
548 patients with asthma. J. Asthma 1995;32:191-197.
- 549 36. Katsumata U, Miura M, Ichinose M, Kimura K, Takahashi T, Inoue H,
550 Takishima T. Oxygen radicals produce airway constriction and
551 hyperresponsiveness in anesthetized cats. Am Rev Respir Dis 1990;
552 141:1158-1162.
- 553 37. Jacobson JM, Michael JR, Jafri MH, Gurtner GH. Antioxidants and
554 antioxidant enzymes protect against pulmonary oxygen toxicity in the
555 rabbit. J Appl Physiol 1990;68:1252-1259.
- 556 38. Kelly FJ. Glutathione: in defence of the lung. Food Chem Toxicol
557 1999;37:963-966.
- 558 39. Cho YS, Lee J, Lee TH, Lee EY, Lee KU, Park JY, Moo HB. Alpha-
559 lipoic acid inhibits airway inflammation and hyperresponsiveness in
560 a mouse model of asthma. J Allergy Clin Immunol 2004;114:429-435.
- 561 40. Corcho I, Hernández F, Reyes N, Carballo AL, Peña O, Reyes T,
562 Yanes L. Cambios del sistema inmune en procesos inflamatorios
563 durante la aplicación de la ozonoterapia. Rev CENIC Cien Biol 1998;
564 29:203-205.
- 565 41. Horiguchi T, Tachikawa S, Handa M, Hanazono K, Kondo R,
566 Ishibashi A, Banno K. Effects of Suplast tosilate on airway inflamma-
567 tion and airway hyperresponsiveness. J Asthma 2001;38:331-336.
- 568 42. Akinarli A, Guc D, Kalayci O, Yigitbas E, Ozon A. Increased
569 interleukin-4 and decreased interferon gamma production in children
570 with asthma: function of atopy or asthma? J Asthma 2002;39:159-165.
- 571 43. Gavett SH, O'Hearn DJ, Li X, Huang SK, Finkelman FD, Wills-
572 Karp M. Interleukin 12 inhibits antigen-induced airway hyper-
573 responsiveness, inflammation, and Th2 cytokine expression in mice.
574 J Exp Med 1995;182:1527-1536.
- 575 44. Munthe-Kaas MC, Carlsen KH, Helms PJ, Gerritsen J, Whyte M,
576 Feijen M, Skinningsrud B, Main M, Kwong GN, Lie BA, Lodrup
577 Carlsen KC, Undlien DE. CTLA-4 polymorphisms in allergy and
578 asthma and the TH1/TH2 paradigm. J Allergy Clin Immunol 2004;
579 114:280-287.
- 580 45. Bocci V, Paulesu L. Studies on biological effects of ozone 1. Induction
581 of interferon gamma on human leucocytes. Hematologica 1990;75:
582 510-515.
- 583 46. Bocci V, Luzzi E, Corradesi F, Paulesu L, Di Stefano A. Studies on
584 the biological effects of ozone: 3. An attempt to define conditions for
585 optimal induction of cytokines. Lymphokine Cytokine Res 1993;12:
586 121-126.
- 587 47. Busse WW, Lemanske RF. Asthma. N Engl J Med 2001;344:350-362.